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Synthesis of exo-3-amino-10-hydroxy-hexacyclo[10.2.1.0^{2,11}.0^{4,10}.0^{4,14}.0^{9,13}]pentadecane-5,7-diene-endo-3-carboxyclic acid and endo-3-amino-10-hydroxyhexacyclo[10.2.1.0^{2,11}.0^{4,10}.0^{4,14}.0^{9,13}]pentadecane-5,7-diene-exo-3-carboxylic acid

Frans J.C. Martins, Hermanus van der Hoven, Agatha M. Viljoen *

Catalysis and Synthesis Research Group, Chemical Resource Beneficiation Focus Area, North-West University, Potchefstroom, 2520, South Africa

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ABSTRACT

Treatment of hexacyclo[7.4.2.0^{1,9}.0^{3,7}.0^{4,14}.0^{6,15}]pentadecane-10,12-diene-2,8-dione with aqueous sodium cyanide produced 2,8-dihydroxy-hexacyclo[7.4.2.0^{1,9}.0^{3,7}.0^{4,14}.0^{6,15}]pentadecane-10,12-diene-2,8-lactam and with sodium cyanide, ammonium chloride and ammonium hydroxide, 2-amino-8-hydroxy-hexacyclo $[7.4.2.0^{1,9}.0^{3,7}.0^{4,14}.0^{6,15}]$ pentadecane-10,12-diene-2,8-lactam was obtained. 10-Hydroxy-hexacyclo $[10.2.1.0^{2.11}.0^{4.10}.0^{4.14}.0^{9.13}]$ pentadecane-5,7-diene-3-one was converted into the corresponding aminonitrile and hexacyclo[10.2.1.0^{2,11}.0^{4,14}.0^{4,14}.0^{9,13}]pentadecane-5,7-diene-10-hydroxy-3-spiro-5'-hydantoin. Treatment of the latter with barium hydroxide produced exo-3-amino-10-hydroxy-hexacyclo $[10.2.1.0^{2.11} \cdot 0^{4.10} \cdot 0^{4.14} \cdot 0^{9.13}]$ pentadecane-5,7-diene-endo-3-carboxylic acid. The isomeric endo-3-amino-10hydroxy-hexacyclo[10.2.1.0^{2,11}.0^{4,10}.0^{4,14}.0^{9,13}]pentadecane-5,7-diene-exo-3-carboxylic acid was obtained from 3 -cyano-3-ureido-hexacyclo $[10.2.1.0^{2,11}.0^{4,10}.0^{4,14}.0^{9,13}]$ pentadecane-5,7-diene-10-ol.

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1. Introduction

It is well known that hydrocarbon moieties promote the transport of drugs across cell membranes and increase their affinity for lipophilic regions in receptor molecules.^{[1](#page-5-0)} The incorporation of cage carbon frameworks into drugs should have the added advantage that metabolic degradation should be retarded by the inherent stability and steric bulk of the cage skeleton thus prolonging the activity and reducing the frequency of drug administration. Areas in which the introduction of an adamantane moiety has been shown to result in longevity of drug action, increased drug potency, speed of action and receptor site specificity include antibacterial activity, \sim anabolic action,^{[3](#page-5-0)} analgesic activity^{[4](#page-5-0)} and antiviral activity.⁵

Amino acids are important building blocks for the synthesis of a large number of biologically important compounds and pharmaceutical drugs. However, a literature survey revealed that the synthesis of α -amino acids with cage carbon frameworks barely received any attention.

As part of a programme that is concerned with the synthesis and chemistry of amino acids with cage structures we have sought to synthesise novel α -amino acids by utilising polycyclic ketones with cage structures as substrates in Strecker reactions. It was previously shown^{[6–8](#page-5-0)} that the dione 1^9 1^9 is not a suitable substrate to obtain

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amino acids via Strecker reactions due to the effect of transannular interactions. We decided to investigate the possible utilisation of 2^{10} 2^{10} 2^{10} (Fig. 1) as starting material for the synthesis of amino acids with rigid polycyclic carbon skeletons.

Figure 1. Cage compounds considered for amino acid synthesis.

2. Results and discussion

Since initial cyanohydrin and subsequent aminonitrile formation is an obvious route for a Strecker reaction, it was decided to investigate the reaction of the dione 2 with sodium cyanide in aqueous medium.

We found that the dione 2 produces the dihydroxy lactam derivative 3 upon treatment with 1 equiv of aqueous sodium cyanide. The dihydroxy lactam 3 is poorly soluble in conventional NMR

Corresponding author. Tel.: $+27$ 018 299 2356; fax: $+27$ 018 299 2350. E-mail address: attie.viljoen@nwu.ac.za (A.M. Viljoen).

solvents. Compound 3 was characterised by conversion into the triacetate 4 (Scheme 1).

A 1 H and 13 C NMR investigation supported the allocated structure of 4. NMR data of 4 are given in [Experimental section](#page-3-0).

Treatment of 2 with an aqueous mixture of 1 equiv sodium cyanide, ammonium chloride and an excess ammonium hydroxide (Strecker reagents) produced the lactam derivative 5. Support for the correctness of the allocated structure of 5 was obtained from a SIMPLE (secondary isotope multiplets from partially labelled entities) experiment.^{[11](#page-5-0)} A proton noise decoupled ¹³C NMR spectrum was recorded in $(CD_3)_2$ SO, which was treated with two drops of a mixture of 60% D₂O and 40% H₂O to impose partial exchange of protons. As expected the quaternary carbon resonance at δ_c 80.58 $(C-8)$ is registered as a double signal due to a β -effect of the OH/OD group where the second signal appears 0.1 ppm upfield. The carbonyl group of the lactam ring experience β - and γ -effects and is registered with signals appearing at 0.1 and 0.02 ppm upfield (effect of NH/ND and OH/OD groups). Amine protons are the most rapidly exchanging and as a result are also the most difficult to inhibit in the neutral state. The carbon resonance at δ_c 75.03 (C-2) suffers therefore only a β -effect from the nearby N–H group (second signal appears 0.1 ppm upfield). Upon forcing complete protonation of the NH2-group in acidic medium a complex spectrum of signals is observed for C-2 due to the slower exchange in the NH \ddagger group.

Lactams can in many cases be hydrolysed to their corresponding amino acids. However, attempted hydrolysis of 3 and 5 under alkaline or acidic conditions only yielded starting material.

A suitable substrate for the synthesis of novel amino acids with cage structures could be derived from the dione 2 by reduction of the latter with zinc and acetic acid whereby the unsaturated ketol 6^{12} 6^{12} 6^{12} is obtained. Treatment of 6 with an aqueous solution of sodium cyanide containing acetic acid produced the cyanohydrin 7 (Scheme 2).

The cyanohydrin 7 and the hydroxy ketone 6 were both converted into the aminonitrile 8 by the treatment of 7 with ammonia or 6 with a mixture of sodium cyanide, ammonium chloride and ammonium hydroxide. These results are remarkable since no aminonitriles could previously¹³ been isolated from similar reactions on the mono keto derivatives 9, 10, 11 and 12 (Fig. 2).

Figure 2. Keto derivatives, which produce labile aminonitriles.

Attempted hydrolysis of the nitrile group of 8 in acidic or alkaline media failed and no amino acid could be obtained.

Hydantoin derivatives are established precursors for the synthesis of amino acids. Treatment of 8 with ammonium carbonate produced the hydantoin 13 as a precipitate. When 6 was treated with a mixture of sodium cyanide, ammonium chloride, ammonium hydroxide and ammonium carbonate the hydantoin 13 again precipitated from the reaction mixture [\(Scheme 3\)](#page-2-0).

The 300 MHz $¹H$ NMR spectrum of 13 shows the presence of</sup> three deuterium exchangeable protons, which are registered at δ_H 7.63, δ _H 7.39 and δ _H 7.26. The resonance at δ _H 7.63 can be assigned to the 3'-nitrogen proton, which is shifted to lower field by the two adjacent carbonyl groups. The resonance at δ_H 7.39 can be associated with the 1'-nitrogen proton and the resonance at $\delta_{\rm H}$ 7.26 with the OH proton. The latter experiences probably a strong deshielding by the diamagnetic anisotropy, which is induced by the nearby carbonyl group (4'-CO) of the hydantoin ring. The calculated^{[14](#page-5-0)} distance between the OH proton and the oxygen atom of the carbonyl group in 13 is 1.695 Å.

In order to discriminate between 13 and 14 as a possible structure for the hydantoin obtained as described above from 8 or 6 ([Scheme 3\)](#page-2-0) an NOE investigation was conducted. The calculated 14 distance between the proton of the 1'-nitrogen atom in 14 and the proton of the OH-group is 1.862 Å. These protons are close enough to experience a nuclear Overhauser effect (NOE).¹⁵ A rotating frame Overhauser (ROESY) experiment¹⁶ showed no NOE effect between the 1'-NH and the OH proton. This observation was verified by NOE-difference and NOESY experiments,^{[15](#page-5-0)} which rules **14** out as a possible structure for the hydantoin.

The isomer 14 should be obtained from cyclisation of the hydantoic acid derivative 16. The latter was prepared ([Scheme 4\)](#page-2-0) from the hydrochloride salt of the aminonitrile 8 by the treatment

with potassium cyanate in glacial acetic acid whereby 15 was obtained. Hydrolysis of 15 with 20% hydrochloric acid at 60 \degree C produced the desired hydantoic acid derivative 16.

The $^1\mathrm{H}$ NMR spectrum of 15 shows the presence of three deuterium exchangeable protons at δ_H 6.86 (OH), δ_H 6.07 (NH₂) and δ_H 5.99 (NH). In a NOESY-experiment nuclear Overhauser effects (NOEs) between the protons resonating at δ_H 6.86 (OH) and δ_H 6.07 (NH₂) as well as between δ_H 6.86 (OH) and δ_H 5.99 (NH) were observed. These observations confirm the close proximity between the OH-group and the NHCONH2-group in 15 and unambiguously confirm the allocated structure of 15.

Attempted cyclisation of 16 with 20% hydrochloric acid at 100 $^{\circ}$ C to produce 14 failed and only starting material was recovered. Normally quantitative conversion of 16 into 14 should be achieved under these reaction conditions.^{[17](#page-5-0)} Thermal cyclisation of 16 could also not be achieved and it was found to be stable up to its melting point.

The hydantoin 13 and the hydantoic acid derivative 16 were both hydrolysed with barium hydroxide^{[18](#page-5-0)} to produce the desired novel amino acids 17 and 19, respectively [\(Scheme 5\)](#page-3-0). As 6 was derived from the meso dione 2 it is obvious that 17 as well as 19 are racemic mixtures of enantiomers.

As expected negative ion FABMS analysis of the amino acids 17 and 19 in sodium hydroxide containing glycerol matrices shows strong molecular ions at m/z 270 ($[M-H]$). Strong molecular ions are obtained at m/z 272 ([M+H]⁺) for both **17** and **19** when hydrochloric acid containing glycerol matrices are used and recording is done in the positive ion mode. The infrared spectra of 17 and 19 are very similar except in the N-H and O-H stretching vibration region where a broad absorption peak is observed around 3427 cm⁻¹ for **19** whereas broad absorption bands at 3350, 3445 and 3552 cm^{-1} are registered for 17.

Further characterisation of the amino acids 17 and 19 was done by selective acetylation of the amino groups by the treatment with acetic anhydride at room temperature. The mono acetates 18 and **20** were obtained in high yield. The EI, MS, 1 H and 13 C NMR spectra supported mono acetylation. The IR spectra of 18 and 20 exhibit significant differences. The ¹H NMR spectra of **18** and **20** recorded in (CD₃)₂SO both exhibit deuterium exchangeable protons at δ_H 7.84 (OH) and δ_H 4.62 (NH) integrating for one proton each. The carboxylic proton of 20 is registered at δ_H 12.19 as a broad signal (δ_H) 12.06–12.32). The carboxylic proton of 18 is recorded at higher field and is observed as a much broader signal around $\delta_{\rm H}$ 12.04 ($\delta_{\rm H}$ 11.40–12.68). A NOESY-experiment performed on 20 revealed the close proximity between the OH and NHAc groups. An NOE effect is observed between the OH proton of C-10 and the NH and $CH₃$ protons of the nearby NHAc group. An NOE effect between the NH proton and the proton resonating at δ_H 2.49 (H-11) is also registered. The latter also shows an NOE effect with the OH proton, which in turn also experiences an NOE effect with the proton resonating at δ_H 2.18 (H-9). The calculated^{[14](#page-5-0)} distance between the OH proton and H-11 in 20 is 2.640 Å and between the NH proton and H-11 3.054 Å. The distance between the NH proton and H-2 is 3.398 Å and no NOE effect could be observed. A NOESY-experiment

performed on 18 showed an NOE effect between the NH proton and the proton resonating at δ_H 2.29 (H-1). The calculated^{[14](#page-5-0)} distance between H-1 and the NH proton of 18 is 2.462 Å. No NOE effect between the NH proton and other methine proton resonances could be observed. The assignment of the resonance at δ_H 2.29 to H-1 in 18 was verified by a COSY-experiment, which shows correlation peaks between the resonance signals of the AB spin system of the methylene protons on C-15 [d, $\delta_{\rm H}$ 1.04 (J_{AB}=10.1 Hz); d, $\delta_{\rm H}$ 1.17 (J_{AB} =10.1 Hz)] and the protons resonating at δ_H 2.29 (H-1) and δ_H 2.25 (H-12).

3. Conclusion

We have succeeded in synthesising racemic mixtures of enantiomers of exo–endo (17) as well as endo–exo (19) amino acid derivatives utilising the meso dione 2. The latter was converted into the ketol 6, which reacted in a remarkable stereoselective manner towards hydantoin (13) and hydantoic acid (16) formation. Hydrolysis of 13 and 16 produced the desired amino acids.

4. Experimental

4.1. General

Infrared spectra (KBr-disc) were recorded on a Nicolet 550 Magna IR spectrometer. EI mass spectra were obtained at 70 eV on a Micromass Autospec-Tof mass spectrometer. ¹H NMR (300 MHz) and ¹³C NMR (75 MHz) spectra were recorded on a Varian Gemini-300 spectrometer. Melting points are uncorrected.

4.2. 2.8-Dihydroxy-hexacyclo[7.4.2.0^{1,9}.0^{3,7}.0^{4,14}.0^{6.15}]pentadecane-10,12-diene-2,8-lactam (3)

A suspension of 2^{10} 2^{10} 2^{10} (1.5 g, 6.64 mmol) in water (60 cm³) was stirred in an ice bath for 10 min. A solution of sodium cyanide (0.4 g, 8.16 mmol) in water (1 cm³) was added drop wise over a period of 3 h and the reaction mixture stirred for a further 15 h at 50 \degree C. Evaporation to dryness on a water bath and extraction on a Soxhlet apparatus with ethyl acetate as extractant produced 3 (1.1 g, 4.09 mmol, mp 240 °C). IR (KBr-disc): ν_{max} 3395, 3360, 3165, 2950, 1680, 1365, 1290, 1180, 1080, 1075, 925 and 720 cm⁻¹. EIMS, m/z 269 $(M⁺)$. Calcd for C₁₆H₁₆NO₃: C, 71.44; H, 5.62; N, 5.21%. Found: C, 71.42; H, 5.60; N, 5.18%.

4.3. Acetylation of 3

A solution of 3 (0.5 g, 1.86 mmol) in acetic acid anhydride (20 cm^3) was refluxed for 2 h. The excess acetic anhydride was decomposed with ice water and the crystalline product 4 (0.38 g, 0.96 mmol, mp 207 °C) filtered off. IR (KBr-disc): $v_{\rm max}$ 3200, 1770, 1740, 1720, 1380, 1340, 1245, 1235, 1220, 1165, 1040 and 720 cm⁻¹. EIMS, m/z 395 (M⁺). Calcd for C₂₂H₂₁NO₆: C, 66.90; H, 5.36; N, 3.55%. Found: C, 66.86; H, 5.33; N, 3.52%. ¹³C NMR (CDCl₃), δ_c 173.91 (s), 169.68 (s), 169.38 (s), 167.45 (s), 124.25 (d), 124.02 (d), 120.62 (d), 119.51 (d), 96.17 (s), 86.64 (s), 53.84 (d), 52.32 (s), 51.87 (s), 51.10 (d), 48.09 (d), 46.98 (d), 45.71 (d), 43.74 (d), 34.22 (t), 27.85 (q), 21.36 (q) and 21.19 (q). ¹H NMR (CDCl₃), δ_H 5.49–5.85 (m, 4×H), 3.36–3.48 (m, 2×H), 2.65–3.01 (m, 4×H), 2.40 (s, 3×H, CH₃), 2.12 (s, 3×H, CH₃), 2.04 (s, 3×H, CH₃), 1.60 (d, J_{AB}=10.6 Hz, ¹/₂ ×CH₂) and 1.29 (d, J_{AB} =10.6 Hz, $\frac{1}{2}$ ×CH₂).

4.4. 2-Amino-8-hydroxy-hexacyclo[7.4.2.0^{1,9}.0^{3,7}.0^{4,14}.0^{6.15}]pentadecane-10,12-diene-2,8-lactam (5)

To an ice cooled mixture of 25% ammonia solution (15 cm³), sodium cyanide (0.4 g, 8.16 mmol) and ammonium chloride (0.4 g, 7.4 mmol) the dione 2 (1 g, 4.42 mmol) was added in one portion and the reaction mixture was stirred for 24 h in a sealed flask immersed in ice water. The reaction mixture was evaporated to dryness and the product $(5, 0.4 \text{ g}, 1.49 \text{ mmol}, \text{mp } 220 \degree \text{C})$ extracted on a Soxhlet apparatus with ethyl acetate as extractant and recrystallised from 1,4-dioxane. IR (KBr-disc): v_{max} 3410, 3395, 3320, 3165, 3000, 1685, 1310, 1240, 1200, 1115, 1060, 920 and 720 cm⁻¹. FABMS (glycerol–HCl) m/z 269 (M+H)⁺. Calcd for C₁₆H₁₇N₂O₂: C, 71.71; H, 6.02; N, 10.45%. Found: C, 71.67; H, 5.99; N, 10.41%. 13C NMR $[(CD_3)_2$ SO], δ_C 173.66 (s), 124.23 (d), 123.64 (d), 123.28 (d), 122.47 (d), 80.58 (s), 75.03 (s), 54.39 (d), 54.18 (d), 53.65 (d), 52.53 (s), 52.19 (s), 51.59 (d), 45.64 (d), 43.23 (d) and 34.59 (t). ¹H NMR $[(CD₃)₂SO]$, δ_{H} 7.79 (s, 1 × H, NH), 5.49–5.77 (m, 4 × H), 4.73 (s, 1 × H, OH), 2.62-2.79 (m, $5\times$ H), 2.17-22.2 (m, 2 \times H), 2.10 (s, 2 \times H, NH₂), 1.52 (d, J_{AB}=9.4 Hz, $\frac{1}{2}$ ×CH₂) and 1.12 (d, J_{AB}=9.4 Hz, $\frac{1}{2}$ ×CH₂).

4.5. 3-Cyano-hexacyclo[10.2.1.0^{2,11}.0^{4,10}.0^{4,14}.0^{9,13}]pentadecane-5,7-diene-3,10-diol (7)

A solution of 6 (0.75 g, 3.3 mmol) in a mixture of water (15 cm^3) and acetic acid (2 cm^3) was stirred in an ice bath and a solution of

sodium cyanide (0.4 g, 8.16 mmol) in water (5 cm 3) was added drop wise. The reaction mixture was stirred for 48 h at room temperature. The product $(7, 0.5 \text{ g}, 1.98 \text{ mmol}, \text{mp } 115 \text{ }^{\circ}\text{C})$ was extracted with dichloromethane and recrystallised from chloroform. IR (KBrdisc): v_{max} 3440, 3235, 3040, 3000, 2920, 2240, 1360, 1285, 1275, 1270, 1140, 1000, 935, 790, 745 and 695 cm⁻¹. EIMS, m/z 253 (M⁺). Calcd for $C_{16}H_{15}NO_2$: C, 75.96; H, 5.98; N, 5.54%. Found: C, 75.91; H, 5.95; N, 5.51%. ¹³C NMR (CDCl₃), δ_c 131.07 (d), 128.79 (d), 126.98 (d), 125.74 (d), 119.97 (s, C=N), 82.57 (s), 81.76 (s), 62.08 (s), 56.29 (d), 56.26 (d), 53.84 (d), 50.99 (d), 50.98 (d), 46.21 (d), 44.21 (d), and 33.32 (t). 1 H NMR (CDCl₃), $\delta_{\rm H}$ 7.27 (s, OH), 5.73–6.13 (m, 4×H), 4.83 (s, 1 \times H, OH), 2.39–2.64 (m, 7 \times H), 1.43 (d, J_{AB}=10.5 Hz, ¹/₂ \times CH₂) and 1.24 (d, J_{AB} =10.5 Hz, $\frac{1}{2}$ ×CH₂).

4.6. 3-Amino-3-cyano-hexacyclo[10.2.1.0^{2,11}.0^{4,10}.0^{4,14}.0^{9,13}]pentadecane-5,7-diene-10-ol (8)

A solution of **7** (1 g, 3.95 mmol) in methanol (20 cm³) was cooled in an ice bath and cooled 25% ammonia (15 cm³) added in small portions. The reaction mixture was stirred for 6 h in a sealed flask immersed in ice water. The crystalline product (8, 0.65 g, 2.58 mmol, mp 170° C) was filtered off and recrystallised from chloroform.

A mixture of $6 \times (0.5 \text{ g}, 2.2 \text{ mmol})$, sodium cyanide $(0.4 \text{ g},$ 8.16 mmol), ammonium chloride (0.4 g, 7.4 mmol) and 25% ammonia (15 cm 3) was stirred in a sealed flask immersed in ice water for 48 h. The precipitated product $(8, 0.39 \text{ g}, 1.55 \text{ mmol}, \text{mp } 170 \degree \text{C})$ was filtered off and recrystallised from chloroform. IR (KBr-disc): v_{max} 3340, 3320, 3300, 2840–3000, 2215, 1290, 1270, 1265, 1140, 860, 840, 730 and 670 cm⁻¹. EIMS, m/z 252 (M⁺). Calcd for C16H16N2O: C, 76.26; H, 6.39; N, 11.10%. Found: C, 76.18; H, 6.36; N, 11.09%. ¹³C NMR (CDCl₃), δ _C 132.42 (d), 129.19 (d), 126.38 (d), 125.17 (d), 122.31 (s), 81.41 (s), 63.90 (s), 63.52 (s), 56.57 (d), 55.74 (d), 55.49 (d), 52.51 (d), 50.39 (d), 46.25 (d), 46.05 (d) and 33.04 (t). 1 H NMR (CDCl₃), δ_H 5.78–6.12 (m, 4×H), 5.28 (br, OH), 2.34–2.72 (m, 7×H), 2.22 (br, NH₂), 1.40 (d, J_{AB} =10.8 Hz, $^{1}/_{2}$ ×CH₂) and 1.23 (d, J_{AB} =10.8 Hz, $^{1}/_{2} \times CH_{2}$).

4.7. Hexacyclo[10.2.1.0^{2,11}.0^{4,10}.0^{4,14}.0^{9,13}]pentadecane-5,7diene-10-hydroxy-3-spiro-5'-hydantoin (13)

A mixture of $\bf{6}$ (0.5 g, 2.2 mmol), sodium cyanide (0.5 g, 10.2 mmol), ammonium chloride (0.5 g, 9.3 mmol), an excess ammonium carbonate (2 g, 20.8 mmol), 25% ammonia (10 cm³) and ethanol (5 cm³) was sealed in a glass tube. The mixture was stirred in a water bath at 60 $\mathrm{^{\circ}C}$ for 24 h. The reaction mixture was allowed to cool to room temperature and the precipitate (13, 0.3 g, 1.01 mmol, mp 310 \degree C dec) was filtered off and recrystallised from ethanol.

A mixture of 8 (0.5 g, 1.98 mmol), ammonium carbonate (2 g, 20.8 mmol), 50 cm³ methanol and 30 cm³ water was stirred at 40 \degree C for 15 h. Ammonium carbonate (2 g, 20.8 mmol) was added and stirring at 40 \degree C continued for 12 h. The reaction mixture was heated on a steam bath to decompose the excess ammonium carbonate. The reaction mixture was allowed to cool to room temperature and the precipitate (13 , 0.32 g, 1.08 mmol, mp 310 \degree C dec) was filtered off and recrystallised from ethanol. Compound 13, IR (KBr-disc): v_{max} 3395, 3260, 3160, 2965, 2920, 1700, 1690, 1615, 1365, 765, 740 and 705 cm⁻¹. EIMS, m/z 296 (M⁺). Calcd for C17H16N2O3: C, 68.98; H, 5.45; N, 9.46%. Found: C, 68.93; H, 5.42; N, 9.42%. ¹³C NMR [(CD₃)₂SO], δ _C 170.18 (s), 152.12 (s), 130.58 (d), 128.06 (d), 127.87 (d), 125.04 (d), 82.05 (s), 72.75 (s), 58.43 (d), 55.49 (d), 54.10 (s), 52.22 (d), 51.01 (d), 46.89 (d), 44.73 (d), 44.22 (d) and 32.06 (t). 1 H NMR [(CD₃)₂SO], $\delta_{\rm H}$ 7.63 (s, 1×H, 3′-NH), 7.39 (s, $1\times$ H, 1'-NH), 7.26 (s, 1 \times H, OH), 5.87–5.94 (m, 1 \times H), 5.72–5.84 (m, 2×H), 5.62 (d, J=10.8 Hz, 1×H), 2.65–2.71 (m, 2×H), 2.48–2.53 (m, 3×H), 2.37–2.39 (m, 2×H), 1.22 (d, J_{AB}=9.7 Hz, ¹/₂ ×CH₂) and 1.10 $(d, J_{AB} = 9.7 \text{ Hz}, \frac{1}{2} \times \text{CH}_2).$

4.8. 3-Cyano-3-ureido-hexacyclo[10.2.1.0^{2,11}.0^{4,10}.0^{4,14}.0^{9,13}]pentadecane-5,7-diene-10-ol (15)

A solution of **8** (0.5 g, 1.98 mmol) in dichloromethane (30 cm³) was saturated with dry hydrogen chloride gas while cooled in ice and stirred for 12 h. The resulting precipitate was collected by filtration (0.52 g, 1.8 mmol) and dissolved in glacial acetic acid (10 cm³). To this solution powdered potassium cyanate (0.3 g, 3.7 mmol) was added slowly in small portions. After the reaction mixture has been warmed for an hour at 60° C, it was poured into five times its volume of cold water. The product (15, 0.47 g, 1.51 mmol, mp 255° C) was filtered off and recrystallised from ethanol. IR (KBr-disc): v_{max} 3460, 3355, 3207, 2975, 2237, 1655, 1615, 1525, 1290, 1260, 1255, 1230, 1120 and 730 cm⁻¹. EIMS, m/z 311 (M⁺). Calcd for C₁₇H₁₇N₃O₂: C, 69.15; H, 5.76; N, 14.24%. Found: C, 69.11; H, 5.73; N, 14.21%. ¹³C NMR $[(CD_3)_2$ SO], δ_C 157.84 (s), 132.25 (d), 127.53 (d), 127.43 (d), 124.50 (d), 120.56 (s), 80.45 (s), 64.12 (s), 61.68 (s), 55.66 (d), 53.84 (d), 53.28 (d), 50.99 (d), 50.81 (d), 45.33 (d), 45.05 (d) and 32.44 (t). 1 H NMR [(CD₃)₂SO], $\delta_{\rm H}$ 6.86 (s, 1 \times H, OH), 6.07 (s, 2×H, NH₂), 5.99 (s, 1×H, NH), 5.93–5.96 (m, 1×H), 5.74–5.85 $(m, 3\times H)$, 2.83–2.86 $(m, 1\times H)$, 2.48–2.53 $(m, 2\times H)$, 2.35–2.38 $(m,$ 2×H), 2.26–2.31 (m, 2×H), 1.33 (d, J_{AB} =10.3 Hz, $\frac{1}{2}$ ×CH₂) and 1.14 $(d, J_{AB} = 10.3 \text{ Hz}, \frac{1}{2} \times \text{CH}_2).$

4.9. Hexacyclo[10.2.1.0^{2,11}.0^{4,10}.0^{4,14}.0^{9,13}]pentadecane-5,7diene-10-hydroxy-3-ureido-3-carboxylic acid (16)

A solution of **8** (0.5 g, 1.98 mmol) in dichloromethane (30 cm³) was saturated with dry hydrogen chloride gas while cooled in ice and stirred for 12 h. The resulting precipitate was collected by filtration (0.52 g, 1.79 mmol) and dissolved in a mixture of glacial acetic acid (9 cm³) and water (5 cm³). After addition of potassium cyanate (1 g, 12.35 mmol) the reaction mixture was refluxed for 2 h. Hydrochloric acid (20%, 4.5 cm³) was added and refluxing continued for 1 h. The reaction mixture was cooled to room temperature and crushed ice added until precipitation started. The product (16, 0.3 g, 1.0 mmol, mp 295 \degree C) was filtered off and recrystallised from ethanol. IR (KBr-disc): v_{max} 3560, 3350, 3060, 2960, 1765, 1725, 1430, 1375, 1285, 1265, 1230, 1080, 790, 758 and 730 cm⁻¹. EIMS, m/z 296 $(M^+$ -H₂O). Calcd for C₁₇H₁₈N₂O₄: C, 64.97; H, 5.73; N, 8.92%. Found: C, 64.91; H, 5.69; N, 8.89%. ¹³C NMR [(CD₃)₂SO], δ _C 176.00 (s), 155.76 (s), 132.10 (d), 127.72 (d), 127.24 (d), 124.68 (d), 80.29 (s), 76.49 (s), 60.68 (s), 54.97 (d), 54.29 (d), 54.04 (d), 53.12 (d), 50.70 (d), 44.27 (d), 42.36 (d) and 42.46 (t). ¹H NMR [(CD₃)₂SO], δ_H 10.83 (br, COOH), 6.95 (s, 1×H, OH), 5.79–5.87 (m, 1×H), 5.81 (s, 1×H, NH), 5.68–5.75 $(m, 2\times H)$, 5.24 (d, J=11.02 Hz), 3.00–3.01 (m, 1×H), 2.40–2.49 (m, $3\times$ H), 2.25–2.32 (m, 2 \times H), 2.19–2.21 (m, 1 \times H), 1.17 (d, J_{AB}=10.6 Hz, $\mathcal{V}_2 \times$ CH₂) and 1.09 (d, J_{AB}=10.6 Hz, ¹ $\mathcal{V}_2 \times$ CH₂).

4.10. Hydrolysis of 15

A mixture of 15 (1.0 g, 3.2 mmol) and 20% hydrochloric acid (10 cm³) was heated at 60 °C for 1 h with stirring. The reaction mixture was evaporated to dryness on a steam bath and the residue washed with cold water and recrystallised from ethanol to produce 16 (0.8 g, 2.5 mmol).

4.11. exo-3-Amino-10-hydroxy-hexacyclo[10.2.1.0^{2,11}.0^{4,10}.0^{4,14}. $0^{9,13}$]pentadecane-5,7-diene-endo-3-carboxylic acid (17)

A mixture of 13 (1.0 g, 3.69 mmol) and a 4% barium hydroxide (60 cm³) solution was stirred in an open beaker in an autoclave for 0.5 h at 180 °C. The reaction mixture was diluted with water

(250 cm³) and an excess of ammonium carbonate (5 g) was added to the reaction mixture and then heated to boiling point with stirring. The hot solution was filtered to remove the insoluble barium carbonate. The pH of the filtrate was adjusted to 6.5 by addition of hydrochloric acid (4 mol dm^{-3}) and evaporated on a steam bath to a small volume ($\pm 5~{\rm cm}^3)$ and the amino acid (**17**, 0.75 g, 2.77 mmol, mp 341 °C dec) filtered off as a colourless solid. IR (KBr-disc): v_{max} 3552, 3445, 3350, 3018, 3003, 2975, 2942, 2894, 2870, 2780, 1620, 1574, 1494, 1331, 1368, 1295, 1242, 1224, 1012, and 729 cm $^{-1}$. FAB (glycerol+HCl matrix) MS (positive ion mode) m/z 272 ($[M+H]^+$). FAB (glycerol+NaOH matrix) MS (negative ion mode) m/z 270 ([M-H]⁻). Calcd for C₁₆H₁₇NO₃: C, 70.91; H, 6.32; N, 5.17%. Found: C, 70.88; H, 6.28; N, 5.15%.

4.12. exo-3-Acetylamino-10-hydroxy-hexacyclo[10.2.1.0^{2,11}.0^{4,10}. $0^{4,14}.0^{9,13}$]pentadecane-5,7-diene-endo-3-carboxylic acid (18)

A solution of 17 (0.5 g, 1.6 mmol) in a 2:1 mixture of acetic anhydride and glacial acetic acid (20 cm 3) was stirred at room temperature for 3 h. Water (100 cm^3) was added and the reaction mixture stirred for 5 h. The mono acetate (18, 0.48 g, 1.52 mmol, mp $218 °C$) precipitated and was filtered off and recrystallised from ethanol. IR (KBr-disc): v_{max} 3464, 3398, 3350, 3018, 2985, 2972, 2904, 1708, 1685, 1629, 1550, 1426, 1283, 1264, 1232, 1159 and 729 cm⁻¹. EIMS, *m*/z 313 (M⁺). Calcd for C₁₈H₁₉NO₄: C, 69.01; H, 6.07; N, 4.47%. Found: C, 68.95; H, 6.04; N, 4.45%. 13C NMR $[(CD₃)₂SO]$, δ_C 172.43 (s), 169.59 (s), 132.76 (d), 130.57 (d), 124.92 (d), 124 .25 (d), 80.45 (s), 71.20 (s), 62.07 (s), 56.25 (d), 54.01 (d), 53.76 (d), 51.45 (d), 51.35 (d), 45.13 (d), 43.90 (d), 32.31 (t) and 22.75 (q). 1 H NMR [(CD₃)₂SO], $\delta_{\rm H}$ 12.04 (br, 1 \times H, COOH), 7.84 (s, 1 \times H, OH), 6.35 (d, J=8.8 Hz, 1×H), 5.56–5.73 (m, 3×H), 4.62 (s, 1×H, NH), 2.42–2.50 (m, $2\times$ H), 2.34–2.37 (m, $1\times$ H), 2.28–2.30 (m, $1\times$ H), 2.23– 2.26 (m, $1\times$ H), 2.16–2.19 (m, $1\times$ H), 2.03–2.05 (m, $1\times$ H), 1.79 (s, $3\times$ H, CH₃), 1.17 (d, J_{AB}=10.1 Hz, $\frac{1}{2}$ ×CH₂) and 1.04 (d, J_{AB}=10.1 Hz, $\frac{1}{2}$ \times CH₂).

4.13. endo-3-Amino-10-hydroxy-hexacyclo[10.2.1.0^{2,11}.0^{4,10}. $0^{4,14}$. $0^{9,13}$]pentadecane-5,7-diene-exo-3-carboxylic acid (19)

A mixture of 16 (1.0 g, 3.69 mmol) and a 4% barium hydroxide (60 cm³) solution was stirred in an open beaker in an autoclave for 1 h at 180° C. The reaction mixture was diluted with water (250 cm 3) and an excess of ammonium carbonate (5 g) was added to the mixture and then heated to boiling point with stirring. The hot solution was filtered to remove the insoluble barium carbonate. The pH of the filtrate was adjusted to 6.5 by addition of hydrochloric acid (4 mol dm $^{-3}$) and evaporated on a steam bath to a small volume (±5 cm 3) and the amino acid (**19**, 0.68 g, 2.51 mmol, mp 350 °C dec) filtered off as a colourless solid. IR (KBr-disc): ν_{max} 3427, 3018, 3003, 2975, 2947, 2894, 2870, 2784, 1614, 1576, 1496, 1381, 1368, 1295, 1242, 1224, 1012 and 729 cm^{-1} . FAB (glycerol+HCl matrix) MS

(positive ion mode) m/z 272 ([M+H]⁺). FAB (glycerol+NaOH matrix) MS (negative ion mode) m/z 270 ([M-H]⁻). Calcd for C₁₆H₁₇NO₃: C, 70.91; H, 6.32; N, 5.17%. Found: C, 70.85; H, 6.30; N, 5.14%.

4.14. endo-3-Acetylamino-10-hydroxy-hexacyclo[10.2.1.0^{2,11}. $0^{4,10}.0^{4,14}.0^{9,13}$ pentadecane-5,7-diene-exo-3-carboxylic acid (20)

A solution of 19 (0.5 g, 1.6 mmol) in a 2:1 mixture of acetic anhydride and glacial acetic acid (20 cm³) was stirred at 60 °C for 2 h. The reaction mixture was cooled to room temperature. Water (100 cm^3) was added and the reaction mixture stirred for 5 h. The mono acetate $(20, 0.3, g 0.96, \text{mmol}, \text{mp } 207, \text{°C})$ precipitated and was filtered off and recrystallised from ethanol. IR (KBr-disc): v_{max} 3350, 3285, 2970, 2950, 2877, 1716, 1595, 1524, 1487, 1422, 1375, 1325, 1290, 1263, 1222 and 789 cm⁻¹. EIMS, m/z 313 (M⁺). Calcd for C18H19NO4: C, 69.01; H, 6.07; N, 4.47%. Found: C, 68.93; H, 6.05; N, 4.43%. ¹³C NMR [(CD₃)₂SO], δ_C 172.41 (s), 169.57 (s), 132.76 (d), 130.55 (d), 124.91 (d), 124 .23 (d), 80.44 (s), 71.19 (s), 62.06 (s), 56.24 (d), 53.99 (d), 53.75 (d), 51.44 (d), 51.34 (d), 45.12 (d), 43.89 (d), 32.29 (t) and 22.74 (q). ¹H NMR [(CD₃)₂SO], δ_H 12.18 (br, 1×H, COOH), 7.84 (s, 1×H, OH), 6.35 (d, J=8.8 Hz, 1×H), 5.56–5.73 (m, $3\times$ H), 4.62 (s, 1 \times H, NH), 2.42–2.49 (m, 2 \times H), 2.34–2.37 (m, 1 \times H), 2.29–2.32 (m, 1×H), 2.24–2.27 (m, 1×H), 2.17–2.19 (m, 1×H), 2.03– 2.05 (m, 1 \times H), 1.79 (s, 3 \times H, CH₃), 1.17 (d, J_{AB}=10.1 Hz, ¹/₂ \times CH₂) and 1.04 (d, J_{AB} =10.1 Hz, $\frac{1}{2}$ ×CH₂).

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